Novel Nanocomposite HNT-TiO$_2$/PVDF Adsorptive Nanofiber Membranes for Arsenic (III) Removal

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1. Introduction

Arsenic is found in earth’s crust in more than 200 different minerals. Leaching of this naturally occurring arsenic has resulted in the persistent contamination of groundwater supplies in several countries around the world including Bangladesh, India and Taiwan $^{[1,2]}$. This has created a significant human health challenge as arsenic is known to cause bladder, lungs, skin, and kidney cancers $^{[3-6]}$. Accordingly, the World Health Organization (WHO) has set the a maximum limit of 10 ppb for arsenic in drinking water $^{[7,8]}$. Various methods have been experienced for arsenic removal including...
chemical and electrochemical treatment, ion exchange, adsorption, and membrane filtration [9-14]. An emerging technology that offers promise in removing arsenic is membrane adsorption (MA). MA combines adsorption and membrane filtration technologies as an efficient method for treating contaminated waters [15-18]. Considering various MA techniques, electrospun nanofiber adsorptive membranes (ENAMs) are of particular interest due to their high permeation rate and adsorption capacity [18-23]. ENAMs are often functionalized via nanoparticles addition to enhance their performance and adsorption capacity. One of such nanomaterials is halloysite nanotube or HNT (Al₂Si₂O₅(OH)₄). HNT is typically 0.5 to 10 nm long, has an inner diameter of 12-15 nm and an outer diameter of 50-60 nm. The outer surface of HNT is mostly composed of SiO₂ and its inner surface is composed of Al₂O₃, hence, it serves as an excellent support material for metal oxides [24-28]. Earlier studies have shown that immobilization of titanium dioxide nanoparticles on HNT improved TiO₂ dispersion and prevented its aggregation [27,29]. Compared to carbon nanotubes, HNTs disperse better in water, are reused more readily, and are cheaper [30,31]. Therefore, immobilization of TiO₂ on the HNT surface offers a promising approach for the removal of arsenic via adsorption [26].

In this study, we reported the preparation and characterization of electrospun nanofibre membranes containing HNT-TiO₂ nanoparticles for the removal of As(III) from contaminated water.

2. Methodology

2.1. Materials

Polyvinylidene fluoride (PVDF® 761) was purchased in pellet form from Kynar polymer. Dimethylacetamide (DMAc, > 99.5%), polyvinylpyrrolidone (PVP), halloysite nanotubes (5-15 nm inside diameter), N-[β-(aminomethyl)-γ-aminopropyl]trimethoxy silane (AEAPTMS), sodium arsenite (Na₃AsO₃), toluene, tetra hydro furane (THF) and acetone (>99.3%) were purchased from Sigma Aldrich. Titanium dioxide P25 (surface area 50±15 m²/g) was purchased from Degussa and Merck supplied sodium hypochlorite (NaClO) and NaOH.

2.2. HNT-TiO₂ synthesis

To immobilize TiO₂ on HNTs surface, 10 g of AEAPTMS was dissolved in 100 mL of toluene via sonication. 10 g TiO₂ and 10 g HNTs were then introduced and the suspension was heated under reflux at 95 ºC for 24 h. TiO₂-HNT was rinsed first with 500 mL THF and then with 500 mL distilled water to remove free TiO₂ and HNTs. TiO₂-HNT produced in this way was separated and dried at 85 ºC for 48 h.

2.3 Dope preparation and spinning procedure

ENAM polymer dope was prepared by dissolving a pre-weighed amount of PVF as pore forming agent and HNTs-TiO₂ in DMAc and sonicated for almost 4 h, followed by mechanical stirring for 12 h to minimize HNTs-TiO₂ agglomeration. PVDF polymer and acetone as non-solvent were then added to the dope, which was also sonicated for an hour to remove air bubbles. The composition of the electro-spinning dope is given in Table 1. The spinning dope was extruded from a 30 mL plastic syringes via a 27-gauge stainless needle (outer diameter 0.4 mm, length 10 mm) at 32 ºC. The voltage was 15 kV, the extrusion flow rate was 1.6 mL/h, and the needle was kept at a fixed distance of 15 cm from the collector. The electrospinning process was conducted at 32 ºC. The collector was wrapped with aluminum foil covered by polyethylene terephthalate (PET) nonwoven support. The ENAMs were pill off collected after 10 h of electrospinning.

Table 1

<table>
<thead>
<tr>
<th>ENAM</th>
<th>PVDF:HNT-TiO₂ (w/w) %</th>
<th>HNT-TiO₂</th>
<th>PVDF</th>
<th>PVP</th>
<th>DMAc+Acetone</th>
</tr>
</thead>
<tbody>
<tr>
<td>N1</td>
<td>1:0</td>
<td>0.0</td>
<td>15.0</td>
<td>2.0</td>
<td>83.0</td>
</tr>
<tr>
<td>N2</td>
<td>1:0.25</td>
<td>3.7</td>
<td>14.4</td>
<td>1.9</td>
<td>80.0</td>
</tr>
<tr>
<td>N3</td>
<td>1:0.50</td>
<td>6.9</td>
<td>13.9</td>
<td>1.8</td>
<td>77.4</td>
</tr>
</tbody>
</table>

2.4. Characterization of nanoparticles and ENAMs

Transmission electron microscope (TEM) Hitachi (HT 7700) was used to observe TiO₂-HNTs after the nanoparticles were dispersed in either methanol or ethanol under sonication for 10 min. To examine the structure of ENAMs, a Hitachi (SU 5000 VP) field emission scanning electron microscope (FESEM) was employed. Samples were mounted on FESEM stubs a carbon tape prior being gold coated (carbon paint used on one corner of samples to ensure good conductivity).

A K-Alpha X-ray photoelectron spectroscopy (XPS) system from Thermo Fisher Scientific (East Grinstead, UK) was used to examine the ENAMs chemical composition. The system used a micro-focused X-ray using a 400µm spot with charge compensation, while a low energy flood of electrons and ions was used as required. Under these conditions, the vacuum pressure within the analytical chamber was about 5×10⁻⁶ mbar. Without charge compensation, the analysis chamber vacuum was in the 10⁻⁹ mbar range. Moreover, X-ray diffraction analysis (XRD) was confirming compatibility of TiO₂-HNTs and PVDF in ENAMs. The video-based optical contact angle measuring system was purchased from Data Physics Instruments (OCA 15EC). The surface hydrophilicity was obtained by membrane placing on the contact angle machine and dropping a 5 µL deionized drop of water to the membrane surface and then capturing by digital camera located inside the contact angle machine which was connected to a computer.

2.5. Adsorption experiment

0.2 g of ENAM sample was added to 200 mL of an aqueous solution of NaAsO₂. The sample was mixed continuously at 400 rpm for 24 h at room temperature using a shaker. Arsenic concentration was determined using an inductive coupled plasma - mass spectrometry (ICP-MS) and inductively coupled plasma - optical emission spectrometry (ICP-OES) analysis provided by Thermo Scientific. The ENAM adsorption capacity was examined in the pH range 2 to 11 using 10 ppm aqueous As(III) solution. The pH was adjusted by adding NaOH solution. In this experiment arsenic ion reduction percentage was defined as:

\[ \% R = \left( \frac{C_i}{C_f} \right) \times 100 \]  

where \( C_i \) and \( C_f \) (in ppm) represent arsenic concentrations at the start of the adsorption experiments and after 24 h, respectively. The adsorption isotherm was established by using aqueous solutions of different arsenic concentrations (100, 150, 200, and 250 ppb). The equilibrium amount of arsenic adsorbed on to ENAMs, \( q_{(\text{mg g}^{-1})} \), was determined from:

\[ q_\text{e} = \frac{V}{M_w} \times (C_i - C_f) \]  

where \( V \) is the solution volume and \( M_w \) is the mass of ENAM sample.

Adsorption kinetics study was carried out under the same conditions as the adsorption isotherm study, except for the initial arsenic concentration, \( C_0 \), of 40 ppm. The amount of adsorbed arsenic at a given time, \( q_t \) (mg g⁻¹), was calculated following a similar approach and using the measured arsenic concentration at time \( t \), i.e. \( C_t \).

2.6. Dynamic adsorption experiment

For filtration experiments, a crossflow ultrafiltration set-up equipped with a permeation cell (effective area 12.56 cm²) was used, as seen in Figure 1. To minimize compaction effects during the pure water permeation test, ENAM was first pressurized to 2 bar and was maintained at this pressure for 30 min. The pressure was subsequently reduced to 0.1 bar and permeate was collected over a period of 30 min. The pure water flux was then calculated from:

\[ J = \frac{V}{A \times t} \]  

where \( V \) is the cumulative volume of permeate obtained during time \( t \) and \( A \) is the surface area of the membrane. To test the removal of As(III), the feed solution containing 103.8 ppb As(III) was pumped from a feed tank to the permeation cell and recycled to the feed tank. Solution volume in the feed tank was maintained at 5.5 L by adding make-up solution from time to time. Filtration experiments for a feed solution was operated for only N3 with low As(III) concentration of pH at 6.7 and pressure equal to 1.0 barg.
3. Result and discussions

3.1. Characterization of adsorbent and ENAMs

TEM characterization was applied for the raw HNTs and modified HNTs-TiO$_2$ to reveal the immobilization of TiO$_2$ on the surface of HNTs. TEM micrographs confirm that TiO$_2$ nanoparticles were deposited on the surface of the HNTs (Figure 2).

Figure 3 shows the X-ray diffraction (XRD) characteristics for (a) immobilized HNTs-TiO$_2$ adsorbent, (b) PVDF membrane without HNTs-TiO$_2$ (N1) and (c) PVDF membrane with HNTs+TiO$_2$ (N3). The XRD patterns of immobilized HNTs-TiO$_2$ adsorbent showed the 3 crystalline spectra at 2$\theta$ of 12.9, 22.5, 27.4, 42.1, 53.7, 61.5 and 71.0. The N3 also possessed similar crystalline patterns at 2$\theta$ of 13.4, 22.9, 28.2, 42.8, 54.3, 62.1 and 71.8. The XRD spectra confirm the presence of HNTs-TiO$_2$ nanoparticles and showed that HNT-TiO$_2$ addition resulted in changing PVDF crystal structure from $\alpha$-crystalline phase in N1 to $\beta$-crystalline phase in N3.

Figure 4 shows FESEM images of the three nanofiber membranes (N1, N2 and N3). As shown, the surfaces of ENAMs N2 and N3 membranes are different from the pristine PVDF nanofiber (N1). The average fiber diameters are below 150 nm for all nanofiber membranes. Also, formation of spider web fibers (thinner than an extended from the trunk fibers) is of importance.

XPS analysis was used to determine the elemental composition of the membranes as listed in Table 2. The results confirm the enhancement of TiO$_2$ in N3 membrane compared to N1 and N2. The increase is expected to enhance the adsorption capacity of N3 compared with N1 and N2, as will be discussed later.

Table 2

<table>
<thead>
<tr>
<th>ENAM</th>
<th>C</th>
<th>F</th>
<th>Ti</th>
<th>Others</th>
</tr>
</thead>
<tbody>
<tr>
<td>N1</td>
<td>60</td>
<td>39</td>
<td>0</td>
<td>1</td>
</tr>
<tr>
<td>N2</td>
<td>53</td>
<td>37</td>
<td>8</td>
<td>2</td>
</tr>
<tr>
<td>N3</td>
<td>52</td>
<td>35</td>
<td>9</td>
<td>4</td>
</tr>
</tbody>
</table>

The contact angle, overall porosity, pure water permeation flux (at 0.1 bar,), and theoretical specific surface area, $S$, of the fiber estimated based on Eq. 4 are listed in Table 3:

$$S = \frac{2\pi rl}{M_m}$$

Where $r$ is the average nanofibers radius, $l$ is the total fibers length, and $M_m$ is the mass of the ENAM sample.

The results indicate that addition of HNTs-TiO$_2$ substantially decreases contact angle of the membranes and turn them from hydrophobic to hydrophilic. This is also reflected in the significant enhancement in pure water flux in N2 and N3 compared to N1. The average fiber diameter also decreases with the addition of HNTs-TiO$_2$, which is probably due to conductivity enhancement of the polymer solution and the ejected filament.

Table 3

<table>
<thead>
<tr>
<th>ENAM</th>
<th>Contact angle ($^\circ$)</th>
<th>Pure water flux (L. m$^{-2}$. h$^{-1}$)</th>
<th>Theoretical specific surface area (m$^2$.g$^{-1}$)</th>
</tr>
</thead>
<tbody>
<tr>
<td>N1</td>
<td>100</td>
<td>973</td>
<td>16.8</td>
</tr>
<tr>
<td>N2</td>
<td>45</td>
<td>4176</td>
<td>17.3</td>
</tr>
<tr>
<td>N3</td>
<td>20</td>
<td>5035</td>
<td>17.9</td>
</tr>
</tbody>
</table>

* Effective area of ENAM was 12.6 cm$^2$.
Fig. 3. XRD spectra of (a) HNTs-TiO$_2$ adsorbent, (b) N1 and (c) N3.

Fig. 4. FESEM images of N1, N2, and N3. Scale bar is 500 nm.
3.2. Adsorption analysis

3.2.1. Effect of pH

The dependency of As(III) adsorption on water pH was examined through a series of experiments using N3 membrane. For this purpose, the adsorption of As(III) was determined at pH values between 2 and 11. The results are shown in Figure 5, which indicate that removal of As(III) was maximum in more acidic region. The adsorption started to decline at higher pH values. That is because arsenate ion has a negative charge enhancing arsenic adsorption by electrostatic attraction. The degree of protonation decreases with increasing pH leading to a decline in arsenic ion adsorption.

3.2.2. Adsorption isotherm

Figure 6 depicts adsorption isotherms for N1, N2, and N3 membranes at pH equal to 3. This pH was selected because – as it was shown in previous Section – the arsenic adsorption was maximum. As expected, the membrane with no HNTs-TiO$_2$ (N1) showed the lowest adsorption capacity. A jump in the capacity was observed when HNTs-TiO$_2$ was added to membranes N2 and N3. However, no linear relationship was observed between amount adsorbed and TiO$_2$ loading. That is, while the TiO$_2$ content of N3 membrane was twice as high as that in N2, the adsorption capacity enhanced only a few percentage points. The highest adsorption capacity was 185.6 µg.g$^{-1}$ for N3 when As(III) concentration in the feed was 250 ppb. The adsorption capacities are expected to slightly lower at pH = 6 to 8. Comparison with data provided in Table 3 shows that the increase in the Langmuir adsorption capacity was proportional to the theoretical specific surface area of the membrane ($R^2 = 0.90$). This suggests that increase in the adsorption capacity by HNTs-TiO$_2$ addition was likely caused by the increase in the specific surface area of the membrane.
Langmuir and Freundlich isotherms, given by Equations (5) and (6), were used to fit the adsorption data:

\[ q_e = \frac{mbC_v}{1 + bC_v} \quad (5) \]

\[ q_e = K_f C_v^{1/n} \quad (6) \]

where \( m \) is the monolayer adsorption capacity (mg g\(^{-1}\)), \( b \) is the Langmuir constant (L mg\(^{-1}\)), \( 1/n \) is the heterogeneity factor, and \( K_f \) represents the Freundlich constant (µg g\(^{-1}\)).

The results of the regression analyses are given in Table 4. According to Table 4, the experimental results exhibited a better fit with the Langmuir model as evident by the correlation coefficients (\( R^2 \)), suggesting that monolayer adsorption was likely a more dominant mechanism in arsenic removal by ENAMs.

### 3.2.3. Adsorption kinetics

Figure 7 shows the variation of As(III) amount adsorbed versus time using N3 ENAM at initial feed concentration of 40 ppm. The adsorption kinetic was obtained using Lagergren’s pseudo-first and second order kinetics given by Equations (7) and (8), respectively.

\[ q_t = q_e (1 - \exp(-K_1t)) \quad (7) \]

\[ q_t = \frac{q_e^2 K_2 t}{1 + q_e K_2 t} \quad (8) \]

Here, \( q_e \) and \( q_t \) (both in mg g\(^{-1}\)) are the equilibrium and instantaneous amount of arsenic, respectively, adsorbed onto ENAMs, \( K_1 \) (h\(^{-1}\)) is the pseudo first order rate constant, and \( K_2 \) (g mg\(^{-1}\).h\(^{-1}\)) is the pseudo second order rate constant. The estimated values for model parameters are listed in Table 5. While the correlation coefficients (\( R^2 \)) were above 0.9 for both kinetic models, a comparison between the actual and theoretical results indicated better fit with the second order kinetics.

### 3.3. Permeation experiments

As shown in Figure 8, arsenic concentration increased in permeate progressively and reached to 10 ppb, which is considered the maximum safe level based on the WHO, after the permeate volume of 1.3 L. Concentration of arsenic in permeate further increased to 18 ppb after 2.0 L of permeate was collected at which point the experiment was stopped and the membrane was regenerated using a mild caustic solution, followed by flushing with DI water. The regenerated membrane produced a similar performance, treating a total volume of 1.3 L of contaminated feed before reaching a permeate concentration of 10 ppb, as shown in Figure 8. The reproducibility was defined as the ratio of the permeate volume when the concentration reached 10 ppb in the second cycle to that in the first cycle. The reproducibility in this study was 93.7%. It should be noted that TiO\(_2\) were leached to the permeate solution negligibly, which was less than 8 ppb during all of experiments. The breakthrough volume, defined as \( C_t/C_o = 0.05 \), for both fresh and regenerated membranes was 0.44 L. However, breakthrough volume is a strong function of the feed flow rate. Earlier studies have shown that permeate concentration increased exponentially with increasing flow rate, as adsorbate compounds could pass through the membrane due to insufficient contact time [32]. Therefore, it is expected that breakthrough volume will decrease as flow rate increases.

### Table 4

<table>
<thead>
<tr>
<th>ENAM</th>
<th>Langmuir model</th>
<th>Freundlich model</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>( m ) (µg g(^{-1}))</td>
<td>( b ) (L mg(^{-1}))</td>
</tr>
<tr>
<td>N1</td>
<td>181</td>
<td>0.06</td>
</tr>
<tr>
<td>N2</td>
<td>189</td>
<td>0.11</td>
</tr>
<tr>
<td>N3</td>
<td>192</td>
<td>0.20</td>
</tr>
</tbody>
</table>

**Fig. 7.** Adsorbed amount versus time (membrane, N3; \( C_i = 40 \) ppm; ENAM = 0.2 g; solution volume = 0.2 L).
Table 5
Estimated parameters for first and second order adsorption kinetic models determined using linear regression.

<table>
<thead>
<tr>
<th>C_i (ppm)</th>
<th>q_e1 (mg.g⁻¹)</th>
<th>q_e2 (mg.g⁻¹)</th>
<th>K_1 (h⁻¹)</th>
<th>R²</th>
<th>q_e2 (mg.g⁻¹)</th>
<th>K_2 (g.mg⁻¹.h⁻¹)</th>
<th>R²</th>
</tr>
</thead>
<tbody>
<tr>
<td>40</td>
<td>31.2</td>
<td>20.2</td>
<td>0.592</td>
<td>0.964</td>
<td>31.9</td>
<td>0.106</td>
<td>0.999</td>
</tr>
</tbody>
</table>

Fig. 8. Results of filtration experiment with low As(III) concentration in the feed (concentration = 103.8 ppb; operating pressure = 1 barg, pH = 3) (A) first cycle (B) After membrane cleaning.

4. Conclusions

The TiO₂-HNT/PVDF ENAMs were fabricated and tested for As(III) adsorption and filtration in this investigation. From the investigational results lead to the following conclusions:

1. The TiO₂-HNTs nanoparticles could be successfully synthesized.
2. TiO₂-HNTs nanoparticles could be embedded in the ENAMs,
3. Incorporated TiO₂-HNTs in ENAMs was significantly increased the hydrophobicity, overall porosity, length of the fibers, and specific surface of the fiber.

4. Furthermore, TiO₂-HNTs was improved the As(III) adsorption significantly.
5. Langmuir isotherm and pseudo second order kinetic model exhibited a better fit with the experimental data than the Freundlich isotherm and pseudo-first order model.
6. Finally, large volumes of clean permeate could be collected by using the crossflow filtration set-up. Thus more, ENAMs also could be used after restoration by cleaning with NaOH solution following by flushing with DI water.
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References


